The Course of Biodegradation of Anionic Detergents by Analyses for Carbon, Methylene Blue Active Elgiforitom Feesacch Serv U. B. Department of Agricu Substance and Sulfate Ion¹ For Official Use

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Abstract

Tallow alcohol sulfates, ether alcohol sulfates and esters of a-sulfo tallow fatty acids were degraded aerobically by sewage microorganisms in a system in which detergent was the sole source of C. Biodegradation was followed by loss of C and methylene blue active substance (MBAS) and formation of SO₄--. Tallow alcohol sulfates were rapidly and completely degraded; ether alcohol sulfates not quite as readily. Reduction in MBAS was rapid for the a-sulfo esters but loss of C and SO₄⁻⁻ formation was incomplete, possibly because of the intermediate formation of a resistant sulfosuccinate. Sodium p-(1methylundecyl) benzenesulfonate (LAS) was the reference standard.

Introduction

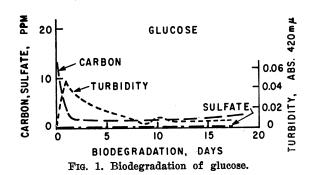
Studies by this laboratory have been reported on the biodegradation of tallow-based detergents in river water (7), in a laboratory scale activated sludge system (3), and in anaerobic digesters (5). An inrestigation of the metabolism of tallow-based detergents by sewage microorganisms (1) has now been extended to give some information on the mechanism of their breakdown under aerobic conditions. The tallow-based detergents were tallow alcohol sulfates, ether alcohol sulfates and esters of a-sulfo fatty acids. Other compounds were included to help relate structure with ease of biodegradation. The individual isomer sodium p-(1-methylundecyl) benzenesulfonate (LAS) was a reference standard.

The Esso Research test (4), with the detergent as the sole source of C, was the basis for a method which permitted measurement of inherent turbidity and analyses for C, methylene blue active substance and SO_4^{--} during $_{
m the}$ (MBAS), biodegradation.

Experimental Procedures

The tests were carried out in widemouth jars containing 3 liters of deionized water, 30 mg of inoculum, nutrient salts free of sulfate and 120 mg of detergent.

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The contents were stirred continuously by magnetic stirrers to insure adequate aeration. Oxygen content, checked with an oxygen meter, was near the saturation point for the test temperature. The activated sludge inoculum was prepared by running sewage samples for one week by a procedure used in confirming tests

Carbon

Total C was determined with a Beckman Carbonaceous Analyzer. Samples were prepared for analysis by centrifuging at 4500 rpm to remove particulate matter, including cells of microorganisms, and the supernatant was purged with N₂ to remove CO₂. In cases where the detergent was not completely soluble the sample was heated to about 80 C to dissolve the detergent, and centrifuged warm.

A Technicon Automatic Analyzer was used. Transmittance was compared with reference standards and expressed as ppm detergent remaining. Samples were used as taken from the aerators without treatment.

Sulfate Ion

Sulfate ion was determined by a turbidimetric method developed in this laboratory (2). The use of isopropyl alcohol instead of absolute ethyl alcohol has been found recently to give a more nearly linear standard curve.

Turbidity

In the method for SO₄-- it was necessary first to measure turbidity in the presence of all reagents except BaCl₂. The values were subtracted from total turbidity before SO₄-- values were computed. The inherent turbidity may give information about the breakdown of detergents and these absorbance values

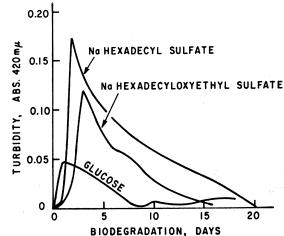
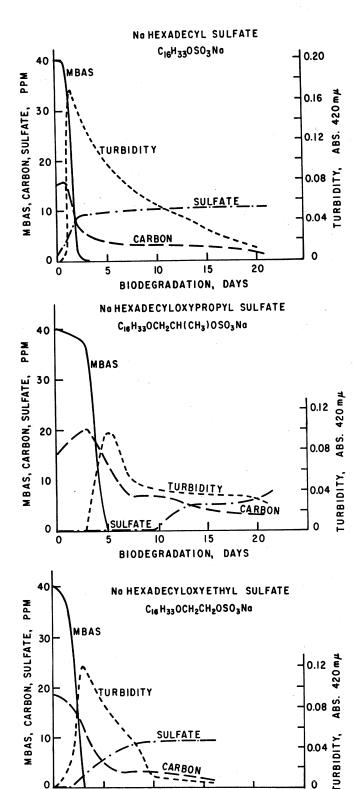


Fig. 2. Comparative turbidities during biodegradation.



BIODEGRADATION, DAYS Fig. 3. Biodegradation of alcohol sulfates and ether alcohol

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were therefore recorded. Hereafter reference to turbidity in this paper is to inherent turbidity.

Results and Discussion

Turbidity may be caused both by the growth of microbial cells and by the formation of insoluble compounds from detergent degradation. In Figure 1 the growth of microbial cells in the biodegradation of

TABLE I Carbon Loss, MBAS Reduction and Sulfate Ion Formed in the Biodegradation of Anionic Detergents

No.	Detergent	Carbon lost, % of theory	SO ₄ formed, % of theory	MBASa Days required for reduction to zero ppm
1	Na hexadecyl sulfate C16H33OSO3Na	94	96	2
2	Na hexadecyloxyethyl sulfate C ₁₆ H ₃₃ OC ₂ H ₄ OSO ₃ Na	95	98	3
3	Na hexadecyloxypropyl sulfate C ₁₆ H ₃₈ OCH ₂ CH (CH ₃)OSO ₃ Na	82	73	5
4	Na hexadecyloxybutyl sulfate C ₁₆ H ₃₃ OCH ₂ CH (C ₂ H ₅)OSO ₃ Na	91	94	5
5	LASb (Ave. of 9 runs) (range)	89 86–95	$\substack{84\\\mathbf{71-92}}$	9
6	Na dodecanesulfonate C ₁₂ H ₂₅ SO ₃ Na	96	100	• 4
7	Na N-methyl-N-sulfoethylpalmitamide C ₁₅ H ₃₁ CON (CH ₃) C ₂ H ₄ SO ₃ Na	94	94	4
8,	Na ₂ sulfoacetate NaO ₃ SCH ₂ CO ₂ Na	100	100	
9	Na dodecyl sulfoacetate C ₁₂ H ₂₅ O ₂ CCH ₂ SO ₃ Na	97	96	3
10	K hexadecyl α-sulfopropionate C ₁₆ H ₃₃ O ₂ CCH(SO ₃ Na)CH ₃	92	91	3
11	Na tetradecyl a-sulfobutyrate C ₁₄ H ₂₉ O ₂ CCH (SO ₃ Na) CH ₂ CH ₃	58	0	5
12	Na hexyl a-sulfopelargonate C ₇ H ₁₅ CH (SO ₃ Na) CO ₂ C ₆ H ₁₃	80	22	9
13	Na dodecafluoroheptyl α-sulfopelargor C7H15CH(SO3Na)CO2CH2(CF2)5CH1	nate 48	3	
14	Na ₂ a-sulfostearate C ₁₆ H ₃₃ CH (SO ₃ Na) CO ₂ Na	70	0	đ
15	Na methyl a-sulfostearate C ₁₆ H ₃₃ CH (SO ₃ Na) CO ₂ CH ₃	87	48	5
16	Na isopropyl α-sulfostearate C16H33CH(SO3Na)CO2CH(CH3)2	74	23	7
17	Na ₂ 2-sulfoethyl α-sulfostearate C ₁₆ H ₂₃ CH (SO ₃ Na) CO ₂ C ₂ H ₄ SO ₃ Na	68	20	7
18	Na sulfosuccinic acid HO ₂ CCH ₂ CH (SO ₃ Na) CO ₂ H	36	0	c
19	Na dioctyl sulfosuccinate C ₈ H ₁₇ O ₂ CCH ₂ CH (SO ₃ Na) CO ₂ C ₈ H ₁₇ °	83	0	10

Methylene blue active substance. Na p-(1-methylundecyl) benzenesulfonate. Not surface active. No color reaction with methylene blue. c Not surface act

glucose reached a maximum value of 0.05 on the absorbance scale in one day and then decreased. On the 16th day a measurable amount of "sulfate" was present. False sulfate values of up to 2 ppm which sometimes appear and later disappear during detergent biodegradation may be due to microbial growth as in the case of glucose.

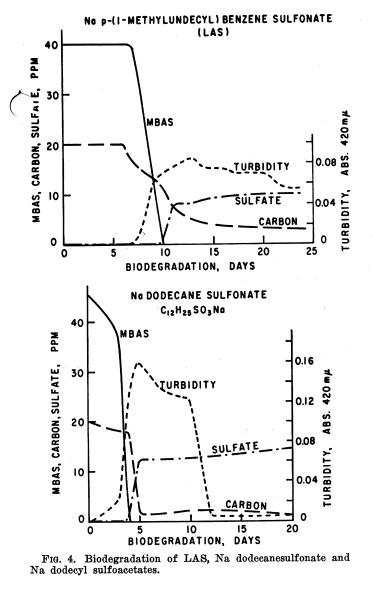
Alcohol sulfates and ether alcohol sulfates, compared with glucose in Figure 2, showed much larger maximum turbidity values possibly due to the separation of insoluble long chain alcohols and ether alcohols.

Alcohol Sulfates and Ether Alcohol Sulfates

Sodium hexadecyl sulfate was rapidly attacked as shown in Figure 3. The MBAS dropped to almost zero in two days and turbidity reached a maximum of 0.175 on the absorbance scale at the same time. Soluble carbon decreased to 27% of the theoretical value and 90% of the theoretical SO₄⁻⁻ was formed. From this point on the reaction slowly approached completion so that after 20 days turbidity was only 0.01 on the absorbance scale, 94% of C had been lost and 96% of theoretical SO₄⁻⁻ was present.

The curves for the ether alcohol sulfates were

similar to that for the alcohol sulfate. Biodegradation was not rapid but proceeded essentially to completion, except that for sodium hexadecyloxypropyl sulfate about 25% of theoretical SO₄-- did not appear an 20% of theoretical C remained. There was no signif icant difference between sodium hexadecyloxyethyl sulfate and sodium octadecyloxyethyl sulfate.



Aliphatic and Aromatic Sulfonates

As shown by Figure 4, biodegradation of sodium dodecanesulfonate was more rapid and complete than for a linear alkylbenzenesulfonate. The value for MBAS dropped to zero in three to four days compared to nine days for LAS; and loss of C and SO₄⁻⁻ formation was nearly quantitative compared to 89% and 84% respectively for LAS. This is true also of other surface active aliphatic sulfonates, not graphed but listed in Table I; specifically sodium N-methyl-N-(2-sulfoethyl) palmitamide (No. 7), sodium dodecyl sulfoacete (No. 9), and potassium hexadecyl asulfopropionate (No. 10).

Esters of α -Sulfo Fatty Acids

With the exception of the fluoro compound (No. 13) all of the a-sulfo esters shown in Figures 5, 6, 7 and Table I, including the sulfosuccinates, were aerobically biodegradable in the sense that MBAS was reduced to zero. Biodegradation was complete for the a-sulfostearates and a-sulfopropionates which showed nearly quantitative loss of C and formation of SO₄⁻⁻. The a-sulfobutyrates and higher homologs of the a-sulfo monocarboxylic acids were less completely degraded, with loss of about 60% to 90% of the theoretical C and formation of 20% to 50% of theoretical SO₄⁻⁻. This may be due to the intermediate

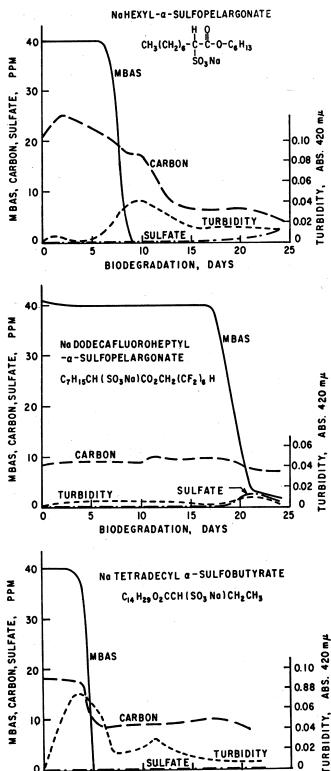


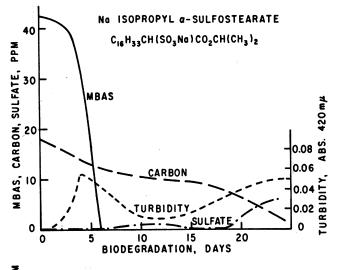
Fig. 5. Biodegradation of α -sulfobutyrates and α -sulfopelargonates.

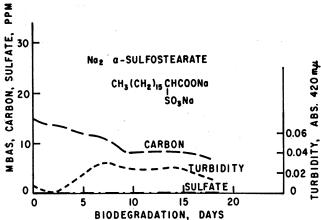
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formation of sulfosuccinates which although biodegradable to the point of loss of surface active properties were resistant to any further decomposition. The sulfo group in this case apparently remains intact without formation of SO₄--. According to this concept, for which there is not much evidence at present, aerobic biodegradation of sulfoacetate, a-sulfopropionates and a-sulfobutyrates may proceed as follows:





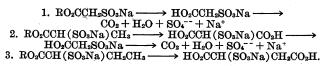


Fig. 6. Biodegradation of a-sulfostearates.

a-Sulfopelargonates, laurates, myristates, palmitates and stearates may degrade through hydrolysis and oxidation to sulfosuccinates which then remain quite resistant to further reaction. Sulfosuccinates are generally considered nontoxic and are useful in the food industry.

a-Sulfo esters such as sodium hexyl a-sulfopelargonate which are wetting agents with the hydrophilic group near the middle of the molecule have

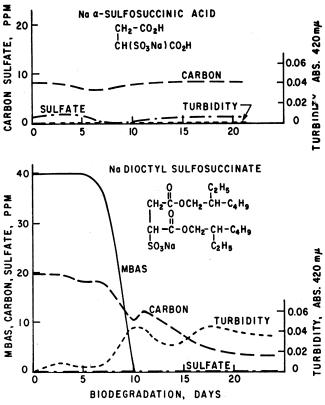


Fig. 7. Biodegradation of sulfosuccinates.

been shown to be less readily biodegradable than a-sulfo esters of the detergent type (7). This is shown again in Table I and in a comparison of Figures 5 and 6. Sodium dodecafluoroheptyl a-sulfopelargonate, also an effective wetting agent may be toxic to microorganisms. Turbidity, as a sign of microbial cell growth remained at a very low level and as a consequence almost no biodegradation occurred until after a period of about 18 days.

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